



Research Article

STUDY ON CHEMICAL CONSTITUENTS OF THE LEAVES OF *STERCULIA FOETIDA* LINN.

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ABSTRACT

Phytochemical data of *Sterculia foetida* Linn. are scarce. The leaves of *Sterculia foetida* Linn. collected in Binh Thuan Province were chemically investigated using multiple chromatographic methods. Three compounds, hesperidin (**1**), kaempferol (**2**) and ursolic acid (**3**) were isolated and elucidated. Their chemical structures were elucidated by comparing their spectroscopic data with those in previous studies. These compounds were found for the first time from the leaves of *Sterculia foetida* Linn. Compound **1** was obtained for the first time in *Sterculia* genus.

Keywords: hesperidin; kaempferol; *Sterculia foetida* Linn.; ursolic acid

1. Introduction

Sterculia foetida Linn. (Sterculiaceae) is grown in tropical areas around the world (Chi, 2002; Peng et al., 2009). The latex of this species can be used as herbal drinks and beverages (Vo, 2002). The *S. foetida* extracts showed antidiabetic, anticancer, antibacterial, anti-inflammatory and analgesic activities (Peng et al., 2009). Flavonoids, triterpenoids, steroids and fatty acids were addressed (Peng et al., 2009; Mujumdar et al., 2000; Anjaneyulu et al., 1981, Kale et al., 2011). Our previous phytochemical studies on this species reported some oleanane-type triterpenoids, quercetin derivatives and phenolic compounds (Pham et al., 2018; Pham et al., 2019). This paper presented the isolation and structural elucidation of three compounds, including hesperidin (**1**), kaempferol (**2**) and ursolic acid (**3**) from *S. foetida* leaves collected in Binh Thuan Province, Vietnam.

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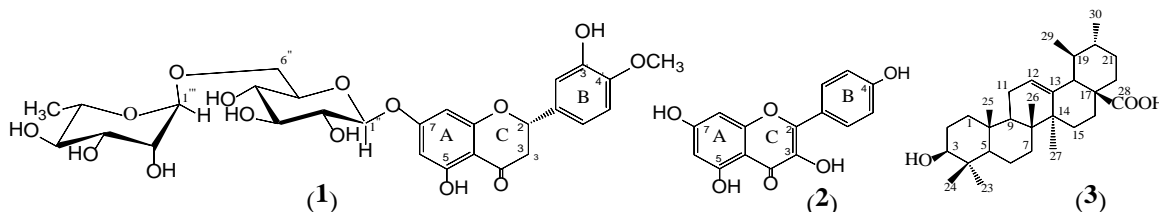


Figure 1. Chemical structures of isolated compounds 1-3

2. Experimental

2.1. General experimental procedures

The NMR spectra were recorded on a Bruker Avance 500 spectrometer (500 MHz for ^1H -NMR and 125 MHz for ^{13}C -NMR) in acetone- d_6 , and DMSO- d_6 solutions. Thin layer chromatography was carried out on silica gel 60 (Merck, 40-63 μm) and spots were visualized by spraying with 10% H_2SO_4 solution, followed by heating.

2.2. Plant material

The leaves of *S. foetida* were collected in Binh Thuan Province, Vietnam in October 2017. Its scientific name was identified by botanist Dr. Dang Van Son, Institute of Tropical Biology. A voucher specimen (No.SFC/TUYEN-1017A) was deposited at the herbarium in the laboratory of Faculty of Environmental Science, Saigon University.

2.3. Extraction and isolation

The powdered leaves of *S. foetida* (12.0 kg) were macerated with ethanol for three times (3 x 40 L) at room temperature. The solvent was removed from the residue under low pressure to obtain the crude extract (1830 g). This crude extract was successively applied to liquid-liquid partition giving hexane (450.5 g), chloroform (650.0 g), ethyl acetate (30.1 g) extracts and the aqueous partition.

The chloroform extract (400.0 g) was applied on a silica gel column chromatography with mobile phase hexane-ethyl acetate (100:0, 75:25, 50:50, 25:75, 0:100, v/v, respectively) and then ethyl acetate-methanol (90:10, 80:20, 70:30, 60:40, 0:100, v/v, respectively) to give eight fractions (SFC.I-SFC.VIII). Fraction SFC.VII (9.2 g) was subjected to a silica gel column chromatography with gradient solvent of chloroform-methanol (80:20, 70:30, 60:40) to yield nine sub-fractions (SFC.VII.1-SFC.VII.9). Sub-fraction SFC.VII.1 (2.3 g) was chromatographed on a silica gel using chloroform-methanol (20:1, 10:1, 5:1) as eluent to give **3** (4.9 mg). Sub-fractions SFC.VII.6 (1.6 g) was rechromatographed on a silica gel eluting with chloroform-methanol (95:5, 90:10, 85:15, 80:20, 75:25) to afford **1** (5.2 mg) and **2** (7.8 mg).

- **Hesperidin (1)** (Lahmer et al., 2015). Yellow amorphous powder. The ^1H -NMR data (500 MHz, DMSO- d_6 , δ ppm, J in Hertz): 12.05 (1H, brs, 5-OH), 6.95 (1H, dd, 8.0, 2.0, H-6'), 6.89 (1H, d, 2.0, H-5'), 6.81 (1H, d, 8.0, H-2'), 6.15 (1H, d, 2.0, H-6), 6.13 (1H, d, 2.0, H-8), 5.49 (1H, dd, 11.0, 5.0, H-2), 4.98 (1H, d, 7.5, H-1''), 4.57 (1H, brs, H-1'''), 3.79 (3H, s, 4'-OCH₃), 3.23 (1H, m, H-3_{ax}), 2.77 (1H, d, 17.0, H-3_{eq}) and 1.09 (3H, d, 5.5,

H-6'''). ^{13}C -NMR (125 MHz, DMSO- d_6): Hesperitin skeleton: 78.1 (C-2), 42.3 (C-3), 197.8 (C-4), 163.0 (C-5), 96.4 (C-6), 165.3 (C-7), 95.6 (C-8), 162.6 (C-9), 103.5 (C-10), 131.1 (C-1'), 114.0 (C-2'), 146.2 (C-3'), 148.2 (C-4'), 112.0 (C-5'), 117.7 (C-6'); D-glucopyranosyl unit: 99.8 (C-1''), 72.0 (C-2''), 75.5 (C-3''), 70.7 (C-4''), 76.3 (C-5'') and 66.0 (C-6''); L-rhamnopyranosyl moiety: 100.1 (C-1'''), 69.7 (C-2'''), 70.4 (C-3'''), 73.1 (C-4'''), 68.3 (C-5''') and 18.1 (C-6'''); and 55.8 (4'-OCH₃).

• **Kaempferol (2)** (Li et al., 2008). Yellow amorphous powder. The ^1H -NMR data (Acetone- d_6 , δ ppm, J in Hertz): 8.14 (2H, d , 8.5, H-2', H-6'), 7.13 (2H, d , 8.5, H-3', H-5'), 6.53 (1H, d , 2.0, H-8) and 6.26 (1H, d , 2.0, H-6). ^{13}C -NMR (125 MHz, Acetone- d_6): 147.0 (C-2), 137.1 (C-3), 176.6 (C-4), 162.3 (C-5), 99.3 (C-6), 166.1 (C-7), 94.5 (C-8), 158.2 (C-9), 104.5 (C-10), 123.6 (C-1'), 130.7 (C-2', C-6'), 116.1 (C-3', C-5') and 160.1 (C-4').

• **Ursolic acid (3)** (Silva et al., 2008). White powder. The ^1H -NMR data (DMSO- d_6 , δ ppm, J in Hertz): 5.14 (1H, t , 4.0;3.5, H-12), 3.20 (1H, dd , 8.0;7.0, H-3), 1.09 (3H, s , H-27), 0.98 (3H, s , H-25), 0.94 (3H, s , H-30), 0.92 (3H, s , H-26), 0.86 (3H, s , H-29), 0.81 (3H, s , H-24) and 0.77 (3H, s , H-23). The ^{13}C -NMR (125 HZ, DMSO- d_6): 38.6 (C-1), 26.9 (C-2), 78.8 (C-3), 36.9 (C-4), 55.2 (C-5), 18.3 (C-6), 31.0 (C-7), 39.4 (C-8), 47.5 (C-9), 36.9 (C-10), 23.2 (C-11), 125.5 (C-12), 138.1 (C-13), 41.9 (C-14), 32.5 (C-15), 22.1 (C-16), 47.7 (C-17), 52.7 (C-18), 30.0 (C-19), 38.8 (C-20), 27.8 (C-21), 36.7 (C-22), 27.9 (C-23), 15.9 (C-24), 15.5 (C-25), 16.8 (C-26), 23.4 (C-27), 180.2 (C-28), 16.9 (C-29) and 21.0 (C-30).

3. Results and discussion

Compound **1** was obtained as a yellow amorphous powder. The ^1H -NMR spectrum of **1** displayed signals of a flavanone skeleton. Its spectrum showed a down field signal at δ 12.05 (1H, brs), indicating the presence of a chelated hydroxyl group at C-5 position. Two *meta*-coupled doublet proton signals at δ_{H} 6.15 (1H, d , 2.0, H-6) and 6.13 (1H, d , 2.0, H-8), each integrated for one proton, were assigned to H-6 and H-8, respectively, of a 5,7-dihydroxy A ring system. The 1',3',4'-trisubstituted B ring system in flavanone skeleton, in addition, was determined by the presence of three aromatic proton signals on ABX system at δ_{H} 6.81 (1H, d , 8.0, H-2'), 6.89 (1H, d , 2.0, H-5') and 6.95 (1H, dd , 8.0, 2.0, H-6'). Furthermore, the flavanone skeleton was identified by the presence of one oxymethine proton at δ_{H} 5.49 (1H, dd , 11.0, 5.0, H-2) and two methylene protons at δ_{H} 3.23 (1H, m , H-3_{ax}) and 2.77 (1H, d , 16.0, H-3_{eq}). These data indicated that **1** was a hesperitin derivative. Moreover, the ^1H -NMR data displayed two anomeric proton signals at δ_{H} 4.98 (1H, d , 7.5, H-1'') and 4.57 (1H, brs , H-1'''), which indicated **1** was a hesperitin diglycoside flavanone. The ^{13}C -NMR spectra were fully supported by the presence of twenty-eight carbon signals, including fifteen carbons of a flavanone skeleton, twelve carbons of two sugar units and one methoxy carbon. The hesperitin flavanone derivative was confirmed by the presence of

fifteen carbons, including one carbonyl carbon at δ_C 197.8 (C-4), twelve aromatic carbon signals from 95.6 ppm to 165.2 ppm, one oxymethine carbon at δ_C 78.1(C-2), and one methylene carbon at δ_C 42.3 (C-3). The β -D-glucopyranosyl unit was demonstrated by the displaying one anomeric proton with the large coupling constant at δ_H 4.98 (1H, *d*, 7.5, H-1'') in the ^1H -NMR spectra and the addition of one anomeric carbon signal at δ_C 99.8 (C-1'') and five oxygenated carbons from 66.0 ppm to 76.3 ppm in the ^{13}C -NMR spectrum. The α -L-rhamnopyranosyl moiety was identified by the presence of one singlet anomeric proton at δ_H 4.57 (1H, *s*, H-1''') along with one anomeric carbon signal at δ_C 100.1 (C-1'''), four oxygenated carbons from 18.1 ppm to 73.1 ppm and one typical L-rhamnopyranosyl methyl carbon at δ_C 18.1 (C-6'''). HMBC correlations from aromatic protons at δ_H 6.81 (1H, *d*, 8.0, H-2'), and 6.95 (1H, *dd*, 8.0, 2.0, H-6') to carbon at δ_C 78.1 (C-2) confirmed the attachment of B benzene ring to C-2. Additionally, the methoxy proton signal at δ_H 3.79 (3H, *s*, 4'-OCH₃) correlated with aromatic carbon at δ_C 148.2 (C-4') in the HMBC spectra which confirmed that the aglycone was hesperetin skeleton. The β -D-glucopyranosyl unit was attached to C-7 of a hesperetin skeleton by the HMBC correlation between the anomeric proton at δ_H 4.98 (1H, *d*, 7.5, H-1'') and oxygenated carbon at δ_C 165.3 (C-7). The α -L-rhamnopyranosyl moiety was linked to C-6 of the β -D-glucopyranosyl unit by the HMBC correlations between the anomeric proton δ_H 4.57 (1H, *s*, H-1''') and oxygenated methylene carbon at δ_C 66.0 (C-6''). The other COSY, HSQC and HMBC correlations were strongly agreed with the assignment. Based on these spectroscopic data and comparison with previous report (Lahmer et al., 2015), the chemical structure of **1** was determined as hesperetin-7-O- $[\alpha$ -L-rhamnopyranosyl-(1 \rightarrow 6)]- β -D-glucopyranoside (Hesperidin). Hesperidin was isolated for the first time from *Sterculia* genus.

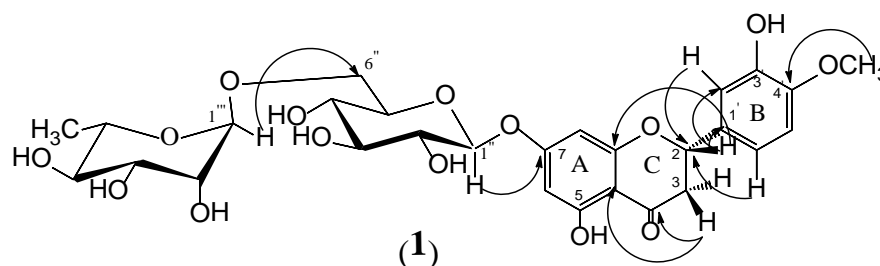


Figure 2. The main HMBC correlations of compound **1**

Compound **2** was obtained as a yellow amorphous powder. Its NMR spectrum showed similar signals of **1**, except for the appearing of one oxygenated aromatic carbon at δ_C 147.0 (C-2), one olefinic carbon at δ_C 137.1 (C-3), and disappearing two sugar units in **2** which was corresponded to a flavonol framework. The *para*-disubstituted benzene ring (ring B) was determined by the displaying of two doublet proton signals with a large coupling constant at δ_H 8.14 (2H, *d*, 8.5 Hz, H-2', H-6') and 7.13 (2H, *d*, 8.5 Hz, H-3', H-5'). These data indicated that **2** was a kaempferol skeleton. The ^{13}C -NMR spectrum were

strongly supported by the displaying of 15 carbons in the down field, comprising one carbonyl carbon at δ_C 176.6 (C-4), and the rest fourteen carbons from 94.5 ppm to 162.3 ppm. The good compatibility between its NMR data and those in the literature (Li et al., 2008) determined the structure of **4** to be kaempferol.

Compound **3** was obtained as a white powder. The NMR data of **3** displayed signals of a triterpenoid skeleton. The $^1\text{H-NMR}$ determined the signals of an olefinic proton at δ_H 5.14 (1H, *t*, 4.0.;3.5, H-12), an oxygenated proton at δ_H 3.20 (1H, *dd*, 8.0;7.0, H-3) and seven methyl proton signals from 0.77 ppm to 1.09 ppm. The $^{13}\text{C-NMR}$ exhibited thirty carbons, including one carboxyl carbon at δ_C 180.2 (C-28), two olefinic carbons at δ_C 125.5 (C-12), and 138.1 (C-13) of a double bond at C-12/C-13 and the rest carbons in ursane skeleton. The HSQC and HMBC spectra determined this suggestion. The comparison NMR data of **3** with those reported in the literature, **3** were assigned as ursolic acid (Silva et al., 2008)

4. Conclusions

From the leaves of *S. foetida* collected in Binh Thuan Province, three compounds, including one diglycoside flavanone hesperidin (**1**), one flavonol kaempferol (**2**) and one ursane-type triterpenoid ursolic acid (**3**) were isolated for the first time. Their chemical structures were determined by using the NMR spectroscopic method as well as comparison with previous studies. Compound **1**, to the best of our knowledge, was isolated from *Sterculia* genus for the first time. Further studies on this species are on the progress.

❖ **Conflict of Interest:** Authors have no conflict of interest to declare.

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**NGHIÊN CỨU THÀNH PHẦN HÓA HỌC CỦA LÁ CÂY TRÔM
STERCULIA FOETIDA LINN.**

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TÓM TẮT

Nghiên cứu hóa thực vật của loài *Sterculia foetida* Linn. chưa được tìm thấy nhiều. Nghiên cứu được thực hiện trên lá cây Trôm *Sterculia foetida* Linn. thu hoạch ở tỉnh Bình Thuận bằng các phương pháp sắc kí khác nhau. Ba hợp chất hesperidin (1), kaempferol (2) và ursolic acid (3) được cô lập và xác định cấu trúc hóa học. Cấu trúc hóa học của các hợp chất được xác định bằng các phương pháp phổ nghiệm đồng thời so sánh với các dữ liệu phổ đã được công bố. Đây là lần đầu tiên các hợp chất này được cô lập từ lá cây Trôm *Sterculia foetida* Linn. Hợp chất 1 lần đầu tiên được biết có hiện diện trong chi *Sterculia*.

Từ khóa: hesperidin; kaempferol; *Sterculia foetida* Linn.; ursolic acid