

## Research Article

**INVESTIGATION OF SOME CULTURE CONDITIONS  
AND NUTRITIONAL SOURCES AFFECTING THE BIOSYNTHESIS  
OF FIBRINOLYTIC ENZYME OF *Bacillus* sp. ES4****Bui Thi Thanh<sup>\*</sup>, Pham Tuan Anh<sup>1,2</sup>, Nguyen Lan Huong<sup>1,2</sup>**<sup>1</sup>*School of Biotechnology and Food Technology, Hanoi University of Science and Technology, Vietnam*<sup>2</sup>*Laboratory of Applied Microbiology, Hanoi University of Science and Technology, Vietnam*<sup>\*</sup>*Corresponding author: Bui Thi Thanh – Email: thanhktb12@gmail.com**Received: March 19, 2023; Revised: March 23, 2023; Accepted: March 24, 2023***ABSTRACT**

*Bacillus* sp. can biosynthesize fibrinolytic enzymes. Culture conditions and nutrient sources are important factors affecting microbial growth and production of the Fibrinolytic enzyme. Therefore, we investigated the culture conditions of the strain *Bacillus* sp. ES4, such as temperature, pH, C nutrient source (glucose, sucrose, glycerol, and maltose), N source (yeast extract, meat content, peptone, and tryptone), metal ions ( $Ca^{2+}$ ,  $Mg^{2+}$ ,  $Mn^{2+}$ ,  $Zn^{2+}$ ,  $K^+$ ,  $Fe^{2+}$ , and  $Cu^{2+}$ ), and time to obtain enzyme product. The results show that for *Bacillus* sp. ES4, the best C source was glucose, peptone and yeast extract.  $Ca^{2+}$  and  $Mg^{2+}$  are nutritional factors that greatly influence the biosynthesis of the fibrinolytic enzyme of strain ES4. At the culture conditions of  $37^{\circ}C$  and  $pH = 6.5$ , the strains for the best enzyme activity at 24 hours and the enzyme activity of the strain when cultured in a 250 ml flask containing 50 ml of medium for the highest enzyme activity at  $450 \pm 25$  FU/ml.

**Keywords:** *Bacillus* sp; culture conditions; enzyme production; fibrinolytic enzyme; optimal nutrient medium

**1. Introduction**

Fibrinolytic enzymes are known to have many advantages in the prevention and treatment of blood clots and many other popular acute and chronic diseases, such as cerebrovascular accidents, myocardial infarction, stroke, and cardiovascular diseases (Zu et al., 2010). It can be eaten directly and has been proven safe during use (Weng et al., 2017). Fibrinolytic enzymes are mainly synthesized by bacterial strains isolated from traditional fermented food sources in Asian countries such as Japan (Sumi, 1987), Korea (Jeong et al., 2009; Yao et al., 2020), China (Wang et al., 2008; Chen et al., 2013; Hu et al., 2019), India (Sharma et al., 2020), Thailand (Chantawannakul et al., 2020), Indonesia (Afifal et al., 2014;

---

**Cite this article as:** Bui Thi Thanh, Pham Tuan Anh, & Nguyen Lan Huong (2023). Investigation of some culture conditions and nutritional sources affecting the biosynthesis of fibrinolytic enzyme of *Bacillus* Sp. ES4. *Ho Chi Minh City University of Education Journal of Science*, 20(3), 479-490.

Yanti, 2018), and Vietnam (Anh et al., 2015; Uyen et al., 2015; Huy et al., 2016; Thu et al., 2020). Isolated strains that can biosynthesize fibrinolytic enzymes include *B. subtilis*, *B. amyloliquefacien*, *B. cereus*, *B. licheniformis*, *Pseudomonas aeruginosa*. The majority are strains of *Bacillus* sp. (Raju et al., 2017; Yogesh et al., 2017; Singh et al., 2018).

The *Bacillus* sp. has high adaptability to the culture medium. However, the enzyme activity biosynthesized from *Bacillus* sp depends on many factors, such as bacterial strains, culture conditions, nutrient composition, and methods. Before that, there have also been studies on optimizing culture conditions and nutrient media of strains to increase enzyme activity, such as those by Liu et al. (2005), Wang et al. (2009), Agrebi et al. (2009), Mahajan et al. (2010), Eldeen et al. (2015), Tuan et al. (2015), Huy et al. (2016), Smitha et al. (2018), and Ju et al. (2019). Therefore, to obtain fibrinolytic enzyme products with high activity, it is necessary to investigate the factors affecting the biosynthesis of enzymes of each strain. In this study, we investigated several factors affecting the production of fibrinolytic enzymes of *Bacillus* sp. ES4 strain, including nutrient sources, culture conditions, and time obtaining enzyme during culture. *Bacillus* sp. ES4 is a mutant strain that has high biosynthesis of the fibrinolytic enzyme. This result is the basis for improving the enzyme activity of *Bacillus* sp. ES4.

## 2. Material and method

### 2.1. Microorganism and media

*Bacillus* sp. ES4 was obtained from the collection at the School of Biotechnology and Food Technology/Hanoi University of Science and Technology (Bui et al., 2022). The strain was stored at -80°C.

The medium for strain activation and propagation is Luria-Bertani (LB) medium consisting of Peptone (10g/l), yeast extract (5g/l), and NaCl (5g/l), solid medium adding agar (15g). /l). The fermentation medium GYP composed of glucose 10 g/L, yeast extract 5 g/L, peptone 5 g/L, NaCl 5 g/L, MgSO<sub>4</sub> 0.25 g/L, and CaCl<sub>2</sub> 0.5 g/L.

### 2.2. Method

#### 2.2.1. Strain activation method

Before use, this strain was activated on an LB agar plate and incubated at 37°C for 24 h. Colonies growing on agar plates were transferred to propagation medium in a 250 ml flask containing 50 ml of LB medium at 37°C for 12-14 h before use.

#### 2.2.2. Culture method to obtain enzymes

After being activated and propagated after 12-14 hours, the strain will be transferred to a 250 ml flask containing 50 ml of GYP medium (OD = 0.2). Culture strain 37°C, shake 150 rpm for 24 hours, enzyme solution obtained after centrifugation to remove cell biomass at 10,000 rpm, temperature 4°C for 10 minutes.

#### 2.2.3. Fibrinolytic enzyme activity assay

The fibrinolytic enzyme activity was determined according to the method described by Thanh et al. (2022). One unit of the fibrinolytic enzyme activity was defined as the amount of enzyme that increased absorbance at 275 nm, equivalent to 1  $\mu\text{g}$  of tyrosine per minute at 37°C.

#### 2.2.4. Determine cell density

Cell growth was monitored by measuring the optical density (OD) of the culture medium at 600 nm using a spectrophotometer (Shimadzu UV-1800, Japan).

#### 2.2.5. Effect of carbon sources on enzyme production

Different strains of bacteria will have different nutritional needs for growth and enzyme production. Therefore, we investigated the effects of the carbon source on the ability to produce fibrinolytic enzymes of *Bacillus* sp. ES4, including glucose, sucrose, maltose, and glycerol. Based on the composition of the GYP medium, the carbon source in the medium was changed after 24 hours of culture shaking at 150 rpm at pH = 7 and a temperature of 37°C. Measure the enzyme activity to select a carbon source for the enzyme-producing strain.

#### 2.2.6. Effect of nitrogen sources on enzyme production

Nitrogen is also the primary source of nutrients for bacterial growth and enzyme biosynthesis. Therefore, to determine a nitrogen source for the *Bacillus* sp. ES4 strain to biosynthesis fibrinolytic enzymes, we investigated the effects of nitrogen sources, including yeast extract, peptone, meat extract, and tryptone. Based on the composition of the GYP medium with a selected carbon source according to Section 2.2.5 and a changed nitrogen source, after 24 hours of culture shaking at 150 rpm at a temperature of 37°C, pH=7.0. Measure the enzyme activity to select a nitrogen source for the enzyme-producing strain.

#### 2.2.7. Effect of metal ions on enzyme production

In addition to the effects of carbon and nitrogen sources, enzyme-producing strains are also affected by metal ions in the culture medium (Cui et al., 2008; Nascimento et al., 2015; Wu et al., 2019 and Salunke et al., 2019). Therefore, we investigated the effects of metal ions, such as  $\text{Ca}^{2+}$ ,  $\text{Mg}^{2+}$ ,  $\text{Mn}^{2+}$ ,  $\text{K}^+$ ,  $\text{Zn}^{2+}$ ,  $\text{Cu}^{2+}$ , and  $\text{Fe}^{2+}$ , on the enzyme biosynthesis ability of the strain, adding metal ions in the form of salts at a concentration of 5 mM ( $\text{CaCl}_2$ ,  $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$ ,  $\text{MnCl}_2 \cdot 4\text{H}_2\text{O}$ ,  $\text{KH}_2\text{PO}_4$ ,  $\text{ZnSO}_4$ ,  $\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$ , and  $\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$ ). Based on a GYP medium with C and N sources as defined in 2.2.5 and 2.2.6, with metal ions in a variable medium. The strain was cultured at 37°C, pH=7 for 24 hours at a shaking speed of 150 rpm, measuring enzyme activity and selecting metal ions for enzyme-producing strains.

#### 2.2.8. Effect of culture temperature on enzyme production

The temperature and pH of the culture medium affect the growth and synthesis of enzyme products of the strain. Therefore, to determine the temperature of *Bacillus* sp. ES4, we investigated the temperature of the culture medium at the values of 30°C, 35°C, 37°C,

40°C, and 45°C with pH 7.0. After incubation, shake at 150 rpm for 24 h. Measure enzyme activity to determine the appropriate culture temperature for the strain.

### 2.2.9. Effect of environmental pH on enzyme production

Based on determining the appropriate temperature for enzyme-producing strains in Section 2.2.8, we continue to investigate the influence of the initial pH of the medium on the enzyme's ability to produce enzymes in the culture medium at pH = 5.5; 6.0; 6.5; 7.0; 7.5 and 8.0. Culture strains were shaken at 150 rpm for 24 hours. Enzyme activity was measured to select the appropriate pH value for the strain.

### 2.2.10. Effect of the culture time to obtain enzymes of the strain

To determine the appropriate time to collect fibrinolytic enzymes of *Bacillus* sp. ES4, we cultured the strain on 50 mL of GYP medium with a determined carbon source, nitrogen source, metal ions, pH, and temperature of the culture medium. After being cultured in flasks for 26 hours and shaken at 150 rpm, samples were taken hourly to measure OD and enzyme activity. The appropriate time to obtain enzyme is at the highest enzyme activity of the strain.

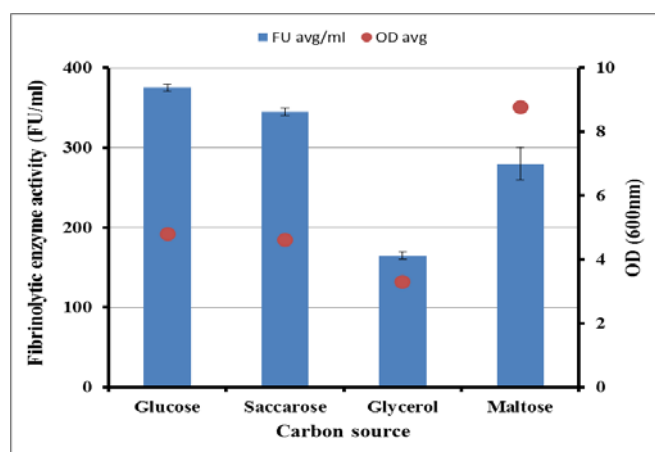
### 2.2.11. Statistical analysis

Process the data with SPSS software, which was used for all statistical analyses.

## 3. Results and discussion

### 3.1. Results on the effects of carbon sources

This strain is grown on media with modified carbon sources: glucose, sucrose, maltose, and glycerol. The effect of the carbon source on the enzyme activity of the strain is shown in Figure 1.



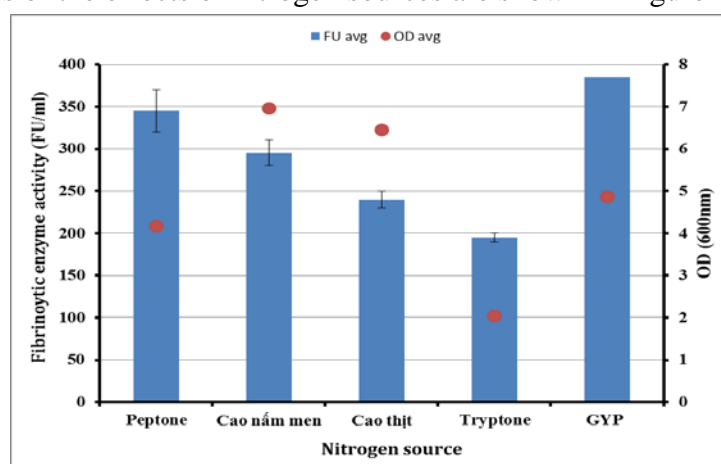
**Figure 1.** Effect of carbon sources on enzyme product

Figure 1 shows that, among the four carbon sources, glucose gave the highest enzyme activity with 375±5 FU/ml, followed by sucrose (345±5 FU/ml), maltose (280±20 FU/ml), and glycerol (165±5 FU/ml). For the glycerol source, the lowest enzyme activity was not equal to half of the glucose source. Therefore, we used the carbon source as glucose to increase the enzyme product of the strain. Some previous studies used glucose as a carbon source to improve the ability to obtain fibrinolytic enzymes (Wang et al., 2009; Kwon et al.,

2011; Tuan et al., 2015). Some studies used glycerol in the culture medium to increase fibrinolytic enzyme production, such as Unrean et al. (2012) and Cui et al. (2020). Xiao et al. (2015) used maltose in the culture medium to increase enzyme production. Vijayaraghave et al. (2017) used sucrose in the culture medium to increase the enzyme product. Thus, carbon sources have different effects on bacterial strains that synthesize fibrinolytic enzymes.

### 3.2. Results on the effects of nitrogen source

Based on the glucose selected, four nitrogen sources, including peptone, yeast extract, meat extract, and tryptone, were investigated for their effects on the enzyme product of the strain. The results of the effects of nitrogen sources are shown in Figure 2.



**Figure 2.** Effect of nitrogen sources on enzyme product

From Figure 2, we see that the nitrogen source was peptone, and yeast extract had higher enzymatic activity than the meat extract and tryptone sources; the highest enzyme activity was  $345 \pm 25$  FU/ml when the strain used a peptone source, and the lowest was a tryptone source reaching  $195 \pm 5$  FU/ml. However, using the combination of two sources of peptone and yeast extract available in the composition of the GYP medium, the enzyme activity of the strain was higher than that of the peptone source ( $385 \pm 5$  FU/ml). This has shown that yeast extract and peptone in the culture medium are more effective. Therefore, to obtain high enzyme activity of the strain, we used peptone and yeast extracts as nitrogen sources in the culture medium. Before, Deepack et al. (2008) and Tuan et al. (2015) used peptone in culture to increase the enzyme activity of strains. Vijayaraghave et al. (2015) used meat extract to increase the enzyme activity of the strain. From the above results, different bacterial strains have different needs for nitrogen sources. Therefore, to obtain high enzyme activity of the strain, we used peptone and yeast extracts as nitrogen sources in the culture medium.

### 3.3. Results on the effects of metal ions

In culturing bacteria, the presence of metal ions in the medium also affects enzyme activity. Figure 3 depicts the effect of metal ions on the results.

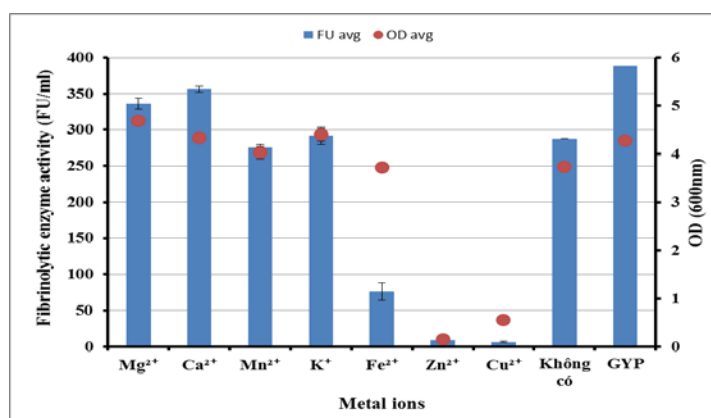


Figure 3. Effect of metal ions on enzyme product

Enzyme activity can increase, decrease, or be inactivated in the media content metal ions. Figure 3 shows that for *Bacillus* sp. ES4, enzyme activity was higher on medium containing metal ions Ca<sup>2+</sup>, Mg<sup>2+</sup>, and K<sup>+</sup> (356±4 FU/ml, 336±8 FU/ml, and 292±12 FU/ml respectively) than on medium without metal ions (288±0 FU/ml). For the four metal ions Mn<sup>2+</sup>, Fe<sup>2+</sup>, Zn<sup>2+</sup>, and Cu<sup>2+</sup>, when added to the culture medium, enzyme activity was lower compared to the non-added medium, indicating that the above metal ions can inhibit enzyme activity. Especially for metal ions Zn<sup>2+</sup> and Cu<sup>2+</sup>, enzyme activity 9.2±1.2 FU/ml and 6.4±0.8 FU/ml, respectively, and enzyme activity has been lost (inactivated). However, when added to the two metal ions Ca<sup>2+</sup> and Mg<sup>2+</sup> in the medium, the enzyme activity of the obtained strain was higher than the medium added one ion, reaching 388±12 FU/ml. This result is similar to the study of Nascimento et al. (2015), which proves that the addition of Ca<sup>2+</sup> and Mg<sup>2+</sup> in the GYP medium is efficient. This result is also consistent with some previously published studies showing that the metal ions Ca<sup>2+</sup> and Mg<sup>2+</sup> affect the biosynthesis of enzymes of the strain (Deepak et al., 2008; Tuan et al., 2015; Xiao et al., 2015; Wu et al., 2019).

### 3.4. Results of the effect of culture temperature

The effect of temperature on the enzyme activity of the strain during culture is shown in Figure 4.

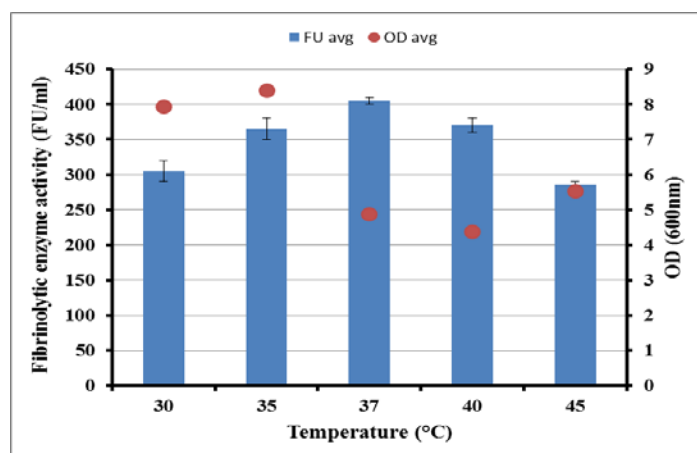
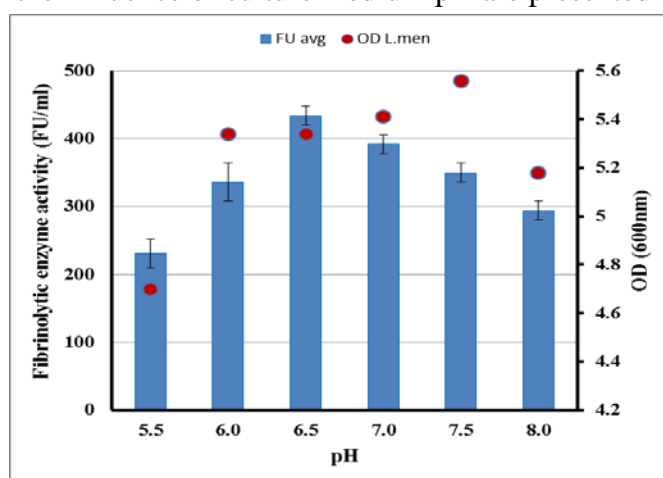


Figure 4. Effect of culture temperature on enzyme product

Figure 4 shows that *Bacillus* sp. ES4 had the highest enzyme activity of  $405 \pm 5$  FU/ml when cultured at  $37^{\circ}\text{C}$  and the lowest enzyme activity at  $45^{\circ}\text{C}$  ( $285 \pm 5$  FU/ml). This shows that, at each temperature value, bacterial strains had different enzyme activities, and when the temperature is high, the enzyme is easy to lose activity. Besides, when the temperature is high, the strain grows slowly, affecting the biosynthesis of the strain's enzyme. Thus, for *Bacillus* sp. ES4, a culture temperature of  $37^{\circ}\text{C}$  was used to obtain the fibrinolytic enzyme.

### 3.5. Results of the effect of pH culture medium

In the culture of bacteria to obtain enzymes, the pH of the initial culture medium also affects the growth and enzyme production of the strain. For *Bacillus* sp. ES4, the results of the investigation on the influence of culture medium pH are presented in Figure 5.

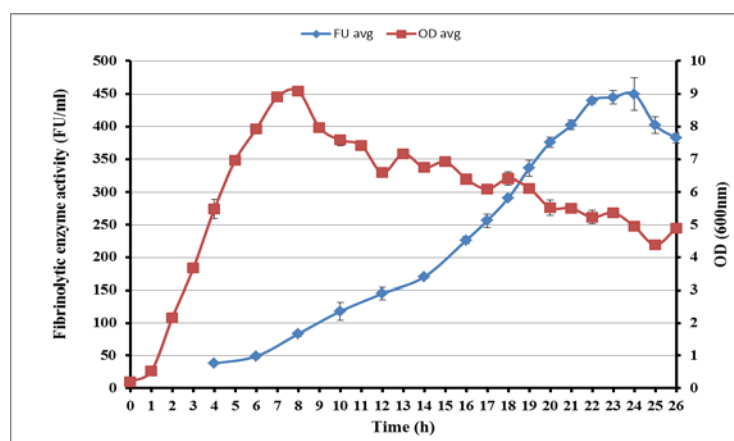


**Figure 5.** Effect of pH culture medium on enzyme product

From Figure 5, it can be seen that, at pH 6.5, the enzyme activity of the strain reached the highest value of  $434 \pm 14$  FU/ml. At pH 7, the enzyme activity of the strain reached  $392 \pm 14$  FU/ml. At pH 5.5, the enzyme activity of the strain was the lowest at  $231 \pm 21$  FU/ml. This shows that, at each pH value, bacterial strains give different enzyme activities. Therefore, the pH of the culture medium affects the enzyme activity of the strain. Strain *Bacillus* sp. ES4 at medium pH 6.5 had the highest enzyme activity, which is the suitable pH for the strain to grow and biosynthesize.

### 3.8. Result of the effect of the culture time to obtain enzymes of the strain

Strains of bacteria grown on different nutrient mediums under different culture conditions will have a different time to obtain enzymes—Strain *Bacillus* sp. ES4 was investigated for the culture time to obtain enzymes; the results are shown in Figure 6.



**Figure 6.** Effect of the culture time to obtain enzymes of the strain

From Figure 6, it can be seen that the bacterial strain developed the strongest biomass at the time of culture from 2 hours to 7 hours, and the OD value reached the highest at 8 hours (OD = 9.085). After 8 hours of culture, strains begin to enter the equilibration phase. Enzyme products were obtained at the end of the equilibration phase; the highest enzymatic activity of the strain was at 24 hours (450±25 FU/ml), followed by enzyme activity at 23 hours and 22 hours (445±10 FU/ml and 440 ±5 FU/ml, respectively). After 24 hours, the enzyme activity of the strain tended to decrease. At 25 hours, the enzyme activity decreased to 402.5±12.5 FU/ml. This can also be due to many reasons. It could be that during the culture process, bacteria can produce substances that are detrimental to the enzyme, that the enzyme is unstable in the culture medium, or bacterial strains can produce inhibitors that lead to a decrease in enzyme activity. The *Bacillus* sp. ES4 strain had the highest enzyme activity of 450±25FU/ml at 24 hours. Therefore, we chose to obtain enzymes of strain cultured in 250-ml flasks at 24 hours for further studies.

#### 4. Conclusion

Strain *Bacillus* sp. ES4 was cultured on a medium with a carbon nutrient source of glucose, a nitrogen source of peptone, and yeast extract, with the addition of metal ions Ca<sup>2+</sup> and Mg<sup>2+</sup> for better biosynthesis of fibrinolytic enzymes compared to other sources.

At a temperature of 37 degrees, the initial pH of the medium was 6.5; *Bacillus* sp. ES4 for the highest fibrinolytic enzyme biosynthesis.

Strain *Bacillus* sp. ES4 has the highest enzyme activity at 24 hours, reaching 450±25 FU/ml in GYP medium at 37°C and pH = 6.5.

❖ **Conflict of Interest:** Authors have no conflict of interest to declare.



## REFERENCES

- Afifah, D. N., Muhammad, S., Dahrul, S., Yanti., & Maggy, T. S. (2014). Isolation and identification of fibrinolytic protease-producing microorganisms from Red Oncom and Gembus, Indonesian fermented soybean cakes. *Malaysian Journal of Microbiology*, 10, 273-279.
- Agrebi, R., Haddar, A., Hajji, M., Frikha, F., Manni, L., Jellouli, K., & Nasri, M. (2009). Fibrinolytic enzymes from a newly isolated marine bacterium *Bacillus subtilis* A26: characterization and statistical media optimization. *Can. J. Microbiol*, 55, 1049-1061. 10.1139/w09-057
- Dinh, B. Q. A., Nguyen, M., Do. N. A. H., & Pham, V. H. (2015). Isolation and Optimization of Growth Condition of *Bacillus sp.* from Fermented Shrimp Paste for High Fibrinolytic Enzyme Production. *Arabian Journal for Science and Engineering*, 40, 23-28. 10.1007/s13369-014-1506-8.
- Bui, T. T., Dam, T. H., Pham, T. A., & Nguyen, L. H. (2022). Enhanced Production of Fibrinolytic Enzyme by *Bacillus sp.* Isolated from Vietnamese Traditional Fermented Soybean (Tuong ban) using Ultraviolet Irradiation and Chemical Mutation. *Int.J.Curr.Microbiol.App.Sci*, 11, 67-80. doi: 10.20546/ijcmas.2022.1105.010
- Chantawannakul, P., Oncharoen, A., Klanbut, K., Chukeatirote, E., & Lumyong, S. (2002). Characterization of proteases of *Bacillus subtilis* strain 38 isolated from traditionally fermented soybean in Northern Thailand. *Science Asia*, 28, 241-245.
- Chen, J. H. B., Zhengbo, H., Qiyi, H., Youjin, H., & Chen, Z. (2013). Isolation and identification of an effective fibrinolytic strain *Bacillus subtilis* FR-33 from the Chinese doufuru and primary analysis of its fibrinolytic enzyme. *African Journal of Microbiology Research*, 7, 2001-2009. 10.5897/AJMR12.282
- Cui. L, Chen, X. C., Jiang, M., Xin, L., & Guijun, Y. (2008). A novel fibrinolytic enzyme from *Cordyceps militaris*, a Chinese traditional medicinal mushroom. *World J Microbiol Biotechnol*, 24, 483-489. DOI 10.1007/s11274-007-9497-1
- Cui, W., Suo, F., Cheng, J., Han, L., Hao, W., & Guo, J. (2018). Stepwise modifications of genetic parts reinforce the secretory production of nattokinase in *Bacillus subtilis*. *Microbial Biotechnology*, 11, doi: 10.1111/1751-7915.13298.
- Deepak, V., Kalishwaralal, K., Ramkumarpanid, S., Venkatesh, B. S., Senthilkumar, S. R., & Sangiliyandi, G. (2008). Optimization of media composition for Nattokinase production by *Bacillus subtilis* using response surface methodology. *Bioresource Technology*, 99, 8170-8174.
- Do. N. A. H., Pham, A. H., & Pham, V. H. (2016). Screening and identification of *Bacillus sp.* isolated from traditional Vietnamese soybean-fermented products for high fibrinolytic enzyme production. *International Food Research Journal*, 23, 326-331.
- Eldeen, K. I., Elrashied, E. E., & Hassan, B. E. (2015). Optimization of Culture Conditions to Enhance Nattokinase Production Using RSM. *American Journal of Microbiological Research*, 3, 165-170. 10.12691/ajmr-3-5-3.
- Fathma S., Narwastu, P., Puspo, E. G., Raymond, R. T., & Maggy, T. S.(2020). Fibrinolytic bacteria of Indonesian fermented soybean: preliminary study on enzyme activity and protein profile. *Food Sci. Technol, Campinas*, 40, 458-465. 10.1590/fst.23919.

- Hu, Y., Yu, D., Zhaoting Wang, Z., Jianjun, H., Tyagi, R., Yunxiang, L. & Yongmei, H. (2019). Purification and characterization of a novel, highly potent fibrinolytic enzyme from *Bacillus subtilis* DC27 screened from Douchi, a traditional Chinese fermented soybean food. *Scientific reports*, 9, 9235-9235. doi: 10.1038/s41598-019-45686-y
- Jeong, W. J., Lee, A. R., Chun, J. Y., Cha, J. H., Song, Y. S., & Kim, J. H. (2009). Properties of cheonggukjang fermented with *Bacillus* strains with high fibrinolytic activities. *Preventive Nutrition and Food Science*, 14, 252-259.
- Ju, S., Cao, Z., Wong, C., Liu, Y., Foda, M. F., & Zhang Z. (2019). Isolation and Optimal Fermentation Condition of the *Bacillus subtilis* Subsp. natto Strain WTC016 for Nattokinase Production. *Fermentation*, 5, p. 92.
- Junguo, L., Chang, T., Zhiya, M., & Huizhou, L. (2005). Optimization of nutritional conditions for nattokinase production by *Bacillus natto* NLSSE using statistical experimental methods. *Process*, 40, 2757-2762. 10.1016/j.procbio.2004.12.025
- Kwon, E. Y., Kim, K. M., Kim, M. K., Lee, I. Y., & Kim, B. S. (2011). Production of nattokinase by high cell density fed-batch culture of *Bacillus subtilis*. *Bioprocess and biosystems engineering*, 34, 789-793.
- Mahajan, P. M., Sagar, V. G., & Smita, S. L. (2010). Production of nattokinase using *Bacillus natto* NRRL 3666: Media optimization, scale up, and kinetic modeling. *Food Science and Biotechnology*, 19, 1593-1603. doi: 10.1007/s10068-010-0226-4
- Naga, R. N., & Divakar, G. (2014). An overview on microbial fibrinolytic protease. *International Journal of Pharmaceutical Sciences and Research*, 5(3), 643-656. doi: 10.13040/IJPSR.0975-8232
- Nascimento, T. P., Amanda, E. S., Porto, C. S., Romero, M. P. B., Galba, M. C. T., & Porto, A. L. F. (2015) Production and characterization of new fibrinolytic protease from *Mucor subitillissimus* UCP 1262 in solid-state fermentation. *Advances in Enzyme Research*, 3, 81-91.
- Nguyen, N. H., & Nguyen, T. H. (2016). Optimization Of The Possibility Synthetic Nattokinase In Soybean Substrates To Orientation Products Development. *International Journal of Pharmaceutical Science Invention*, 5, 35-41.
- Nguyen, Q. U., Nguyen, H. N., Phan, T. H., & Nguyen, H. M. Q (2015). Buoc dau nghien cuu nattokinase cua chung vi khuan *Bacillus* sp. phan lap tu nem chua [The first research on nattokinase of *Bacillus* sp. isolated from Nem Chua]. *Journal of Biology*, 37, 129-133. 10.15625/0866-7160/v37n1se
- Nguyen, A. T., Dinh, T. H. T., Tran, T. M. T, & Nguyen, T. H. (2015). Determination the optimum fermentation in obtaining nattokinase by *Bacillus subtilis* natto. *International Journal of Innovation and Applied Studies*, 13, 663-668.
- Nguyen, T. A. T., Nguyen, T. M. K., Nguyen, D. H., Nguyen, Q. D. T., & Nguyen, H. L. (2020). Characterizations and fibrinolytic activity of serine protease from *Bacillus subtilis* C10. *Current Pharmaceutical Biotechnology*, 21, 110-116.
- Ping, X., Yao, S. P., Liu, J. F., Ying, M., & Wang, Y. P. (2015). Enhanced Production of Fibrinolytic Enzyme from *Bacillus amyloliquefaciens* CGMCC 7380 Using Broad Bean as Substrate. *Advance Journal of Food Science and Technology*, 9, 832-839. doi: 10.19026/ajfst.9.1639

- Salunke., A. S., & Arun, S. K. (2019). Data on isolation and purification of fibrinolytic enzyme from *Pseudomonas baetica* SUHU25. *Data in Brief*, 26, doi.org/10.1016/j.dib.2019.104369
- Sharma. D., Shekhar, S., Kumar, A., & Godheja, J. (2020). Isolation, characterization, production and purification of fibrinolytic enzyme nattokinase from *Bacillus subtilis*. *IJPSR*, 31, 1768-1776. doi.org/10.13040/IJPSR.0975-8232
- Singh, P., Negi, R., Sharma, V., Rani, A., Pallavi, & Prasad, R. (2018). Production of fibrinolytic enzyme (Nattokinase) from *Bacillus sp.* *Indo American Journal of Pharmaceutical Sciences*, 5, 379-383. doi.org/10.5281/zenodo.1155529
- Smitha, K. V., & Pradeep, B. V. (2018). Optimization of Physical and Cultural Conditions of Fibrinolytic Enzyme from *Bacillus altitudinis* S-CSR 0020. *Journal of Pure and Applied Microbiology*, 12, 343-354. doi: 10.22207/JPAM.12.1.40
- Sumi, H., Hamada, H., Tsushima, H., Mihara, H., & Muraki, H. (1987). A novel fibrinolytic enzyme (nattokinase) in the vegetable cheese natto; a typical and popular soybean food in the Japanese diet *Experientia.*, 43, 1110-1111. http:doi:10.1007/BF01956052
- Unrean. P., Nguyen, N. H. A., Visessanguan, W. & Kitsubun, P. (2012). Improvement of nattokinase production by *Bacillus subtilis* using an optimal feed strategy in fed-batch fermentation. *KKU Res. J*, 17, 769-777.
- Vijayaraghavan, P. P. R., Samuel, G. P. V., Arumugaperumal, A., Naif, A. A. D., Mariadhas, V. A., Oh, Y. K., & Kim, Y. O. (2017). Novel Sequential Screening and Enhanced Production of Fibrinolytic Enzyme by *Bacillus sp.* IND12 Using Response Surface Methodology in Solid-State Fermentation. *BioMed Research International*, 2017, 1-13. doi: 10.1155/2017/3909657
- Vijayaraghavan. P., & Samuel, G. P. V. (2014). Statistical Optimization of Fibrinolytic Enzyme Production Using Agroresidues by *Bacillus cereus* IND1 and Its Thrombolytic Activity In Vitro. *BioMed research international*, 2014, 725064. doi: 10.1155/2014/725064
- Wang. J. K, Hua, H. C., & Ching, S. H. (2009). Optimization of the medium components by statistical experimental methods to enhance Nattokinase activity. *Fooyin Journal of Health Sciences*, 1, 2127. 10.1016/S1877-8607(09)60004-7
- Wang. S. H, Zhang, C., Yang, Y. L., & Diao, M. (2008). Screening of a high fibrinolytic enzyme producing strain and characterization of the fibrinolytic enzyme produced from *Bacillus subtilis* LD-8547. *World Journal of Microbiology and Biotechnology*, 24, 475-482. 10.1007/s11274-007-9496-2
- Weng, Y., Yao, J., Sparks, S., & Wang, K. Y. (2017). Nattokinase: An Oral Antithrombotic Agent for the Prevention of Cardiovascular Disease. *International Journal of Molecular Sciences*, 18, 523. doi: 10.3390/ijms18030523
- Wu, R., Chen, G., Pan, S., Zeng, J., & Liang, Z. (2019). Cost-effective fibrinolytic enzyme production by *Bacillus subtilis* WR350 using medium supplemented with corn steep powder and sucrose. *Scientific Reports*, 9.
- Yanti (2018). Screening, Purification, and Characterization of Fibrinolytic Enzyme-Producing Bacteria from Indonesian Fermented Foods. *Scholars Academic Journal of Biosciences*, 6, 598-605. http:doi.10.21276/sajb.2018.6.8.7
- Yogesh, D., & Halami, P. M. (2017). Fibrinolytic enzymes of *Bacillus spp.*: An overview. *International Food Research Journal*, 24, 35-47.

- Zhuang, Y., Yu, M., Le, H. G., Lee, S. J., Hye, S. J., Ji, Y. Y., Diana, N. A., & Kim, J. H. (2020). Isolation of 2 *Bacillus* Strains with Strong Fibrinolytic Activities from Kimchi. *Microbiol. Biotechnol. Lett.*, 48, 439-446. 10.48022/mb1.2003.03008
- Zu, X. Y., Zhang, Z. Y., Yang, Y. N., Che, H. T., Zhang, G. H., & Li, J. (2010). Thrombolytic activities of nattokinase extracted from *Bacillus subtilis* fermented soybean curd residues. *Int. J. Biol.* 2, 120-125.

**KHẢO SÁT ĐIỀU KIỆN NUÔI CẤY VÀ NGUỒN DINH DƯỠNG  
ẢNH HƯỞNG ĐẾN QUÁ TRÌNH SINH TỔNG HỢP ENZYME TIÊU SỢI HUYẾT  
CỦA CHỦNG *Bacillus* sp. ES4**

**Bùi Thị Thanh<sup>1\*</sup>, Phạm Tuấn Anh<sup>1,2</sup>, Nguyễn Lan Hương<sup>1,2</sup>**

<sup>1</sup>*Viện Công nghệ Sinh học và Công nghệ Thực phẩm, Đại học Bách khoa Hà Nội, Việt Nam*

<sup>2</sup>*Phòng thí nghiệm Vi sinh Ứng dụng, Đại học Bách khoa Hà Nội, Việt Nam*

*\*Tác giả liên hệ: Bùi Thị Thanh – Email: thanhktb12@gmail.com*

*Ngày nhận bài: 19-3-2023; ngày nhận bài sửa: 23-3-2023; ngày duyệt đăng: 24-3-2023*

**TÓM TẮT**

*Vi khuẩn Bacillus sp. có khả năng sinh tổng hợp enzym tiêu sợi huyết. Điều kiện nuôi cấy và nguồn dinh dưỡng là những yếu tố quan trọng ảnh hưởng đến sự phát triển và sản xuất enzyme của chủng. Vì vậy, trong nghiên cứu này, chúng tôi khảo sát điều kiện nuôi cấy của chủng Bacillus sp. ES4 như nhiệt độ, pH, nguồn dinh dưỡng C (glucose, sucrose, glycerol và maltose), nguồn dinh dưỡng N (cao nấm men, cao thịt, peptone và tryptone), các ion kim loại (Ca<sup>2+</sup>, Mg<sup>2+</sup>, Mn<sup>2+</sup>, Zn<sup>2+</sup>, K<sup>+</sup>, Fe<sup>2+</sup> và Cu<sup>2+</sup>) và thời gian thu nhận sản phẩm enzym. Kết quả cho thấy đối với Bacillus sp. ES4, nguồn C tốt nhất là glucose; nguồn N tốt nhất là pepton và cao nấm men; Ca<sup>2+</sup> và Mg<sup>2+</sup> là những yếu tố dinh dưỡng có ảnh hưởng lớn đến quá trình sinh tổng hợp enzym tiêu sợi huyết của chủng. Ở điều kiện nuôi cấy 37<sup>0</sup>C và pH = 6,5, chủng cho hoạt độ enzym tốt nhất ở 24 giờ, chủng nuôi cấy trong bình tam giác 250ml chứa 50ml môi trường cho hoạt độ enzym cao nhất đạt 450±25. FU/ml.*

**Từ khóa:** *Bacillus sp.*; điều kiện nuôi cấy; sản xuất enzyme; enzyme tiêu sợi huyết; môi trường dinh dưỡng tối ưu